# KAEMPFEROL AND QUERCETIN GLYCOSIDES FROM RUBUS IDAEUS L. LEAVES

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**Abstract:** Quercetin 3–0– $\beta$ –D–glucoside (I), quercetin and kaempferol 3–0– $\beta$ –D–galactosides (II, III), kaempferol 3–0– $\beta$ –D–coumaroyl)–glucoside (tiliroside) (V) and methyl gallate (VI) were isolated from *Rubus idaeus L.* subspecies culture of Norna leaves and fully characterized.

Keywords: Rubus idaeus, Rosaceae, kaempferol and quercetin glycosides, methyl gallate.

Our previous chemical investigation of the flavonoid compounds present in the leaves of *Rubus idaeus L.*, subspecies culture of Norna, dealt with: quercetin, kaempferol, quercetin  $3-0-\alpha-L$ -arabinopyranoside, kaempferol  $3-0-\beta-D$ -glucuronide and ellagic acid, which were isolated and fully characterized (1).

As a continuation of that work, the isolation and characterization of a few next flavonoid compounds and methyl gallate present in the same source is described.

## **EXPERIMENTAL**

**Plant material** and extraction have been described previously (1)

Chromatography

PC: Whatman No. 1 Solvent systems: S–1 n–BuOH – HOAc –  $H_2O$  (4:1:5) upper phase; S–2 15% HOAc; S–3 HOAc – conc HCl –  $H_2O$  (30:3:10); S–4  $C_0H_6$ – HOAc –  $H_2O$  (6:7:3) upper phase.

TLC: plates cellulose precoated (Merck) – TLC<sub>c</sub>; plates silica gel 60 precoated (Merck) – TLC<sub>g</sub> Solvent systems: S–5 EtOAc– 85% HCO-OH –  $\dot{H}_2O$  (18:1:1); S–6 n–BuOH–pyridine– $\dot{H}_2O$  (6:4:3). CC was achieved on polyamid (Roth). Solvent systems: S–7  $\dot{H}_2O$ –MeOH; S–8  $\dot{C}_6H_6$ –MeOH (both steep gradient).

The spots of flavonoids were visualized under UV light (366 nm) before and after spraying with 2%AlCl<sub>3</sub> in MeOH and 0.5% NA–reagent in MeOH. Sugars were visualised by spraying with aniline phthalate and heating at 105°C.

#### **Isolation**

Compounds V (40 mg) and VI (50 mg) have been obtained from the etheral extract after

column chromatography separation (polyamide, S-7).

Compound V was eluted with 60% aqueous MeOH as a solvent system. Compound VI was eluated with water and further purified on a polyamide column (S-8).

Compounds I (45 mg), II (140 mg), III (15 mg) and IV (40 mg) have been obtained from ethyl acetate extract after separation on the polyamide column (S-7).

Coumpound **I**, **II** and **III** were eluted with 40% aqueous MeOH (as the first fractions) and further purified on the polyamide column (S-8).

Compounds IV and some amount of quercetin  $3-0-\alpha$ –L—arabinopyranoside have been found in further fractions eluted with 40% aqueous MeOH and were further purified on the polyamide column (S-8).

# Identification

Melting points (m.p.) uncorrected were determined on a Boetius apparatus.

Flavonoids were identified by chromatographic analysis of acid hydrolysates and by spectroscopic methods. Total acid hydrolysis was carried out with 5% HCl for 3 h under reflux. EtOAc extracts of hydrolysates were analysed for aglycones (PC, S–3) and H<sub>2</sub>O residues for sugars (TLC<sub>c</sub>, S–6). The UV spectra (Unicam SP800) of flavonoids were recorded in (a) MeOH, also after the addition (b) NaOMe, (c) AlCl<sub>3</sub>, (d) AlCl<sub>3</sub>/HCl, (e) NaOAc, (f) NaOAc/H<sub>3</sub>BO<sub>3</sub> according to Mabry et al. (2).

IR spectra were recorded on an ATI – Mattson FTIR apparatus (compounds V, VI). <sup>1</sup>H NMR spectra were recorded on a Bruker MSL 300

(compounds **I** – **IV**) and DRX 500 (compounds **V**, **VI**), <sup>13</sup>C NMR on DRX 500 (compounds **IV**, **V**); <sup>1</sup>H NMR at 300.13 MHz and 500.13 MHz, <sup>13</sup>C NMR at 125.75 MHz respectively (TMS as internal standard). Additionally, for compound **V** 2D NMR – HMQC the spectrum was recorded on a DRX 500 apparatus.

**Quercetin 3–0–\beta–D–glucopyranoside** (I), yellow needles (MeOH), m.p. 186–189°C.

Rf:  $TLC_g$  S-5, 0.52. UV  $\lambda$  max: a) 255, (268), (300), 358; b) 270, 325, 410; c) 270, (307), (330), 438; d) 265, (300), (358), 402; e) 271, 322, 384; f) 260, (298), 382.

 $^{1}$ H NMR (DMSO – d<sub>6</sub>) δ ppm: 12.63 (s, 1H, OH–5), 7.58 (dd J<sub>1</sub>=2.1 Hz, J<sub>2</sub>=8.1 Hz, 1H, H–6'), 7.57 (d, J=2.1 Hz, 1H, H–2'), 6.83 (d, J=8.5 Hz, 1H, H–5'), 6.39 (d, J=2.0 Hz, 1H, H–8), 6.19 (d, J=2.0 Hz, 1H, H–6), 5.46 (d, J=7.5 Hz, 1H, H–1''), 3.59 – 3.07 (m, 6H of glucose + H<sub>2</sub>O).

#### Acid hydrolysis: quercetin, glucose

**Quercetin 3–0–β–D–galactopyranoside** (**II**), yellow needles (MeOH), m.p. 247–248 °C.

Rf: TLC<sub>g</sub>, S–5, 0.47. UV λ max: a) 257, (270), (297), 360; b) 271, 330, 410; c) 274, (307), (331), 440; d) 267, (300), (360), 406; e) 272, 322, 386; f) 261, (300), 381.

 $^{1}$ H NMR (DMSO–d<sub>6</sub>) δ ppm: 12.63(s, 1H, OH–5), 7.66 (dd, J<sub>1</sub>=2.2 Hz, J<sub>2</sub>=8.5 Hz, 1H, H–6'), 7.52 (d, J=2.2 Hz, 1H, H–2'), 6.81 (d, J=8.5 Hz, 1H, H–5'), 6.40 (d, J=2.0 Hz, 1H, H–8), 6.19 (d, J=2.0 Hz, 1H, H–6), 5.37 (d, J=7.7 Hz, 1H, H–1''), 3.77 – 3.16 (m, 6H of galactose + H<sub>2</sub>0).

# Acid hydrolysis: quercetin, galactose

**Kaempferol 3–0–β–D–galactopyranoside** (III), pale yellow needles – (MeOH), m.p.  $255-257^{\circ}$ C.

Rf: TLC<sub>g</sub>, S–5, 0.48. UV λ max: a) 263, (293), (319), 349; b) 270, (325), 405; c) 272, (303), 345, 400; d) 272, (301), 343, 400; e) 270, (300), 370; f) 265, (291), (320), 350.

 $^{1}$ H NMR (DMSO–d<sub>6</sub>) δ ppm: 12.61 (s, 1H, OH–5), 8.06 (d, J=8.9 Hz, 2H, H–2', H–6'), 6.86 (d, J=8.9 Hz, 2H, H–3', H–5'), 6.43 (d, J=2.0 Hz, 1H, H–8), 6.20 (d, J=2.0 Hz, 1H, H–6), 5.40 (d, J=7.6 Hz, 1H, H–1''), 3.66 – 3.27 (m, 6H of galactose + H<sub>2</sub>0).

## Acid hydrolysis: kaempferol, galactose

**Kaempferol 3–0–α–L–arabinopyranoside** (**IV**), pale yellow needles (MeOH), m.p. 199–202°C. Rf: TLC<sub>g</sub>, S–5, 0.60. UV  $\lambda$  max: a) 265, (293), (322), 350; b) 274, (325), 404; c) 273, (303), 349, 400; d) 273, (304), 347, 400; e) 274, (302), 369; f) 268, (294), (321), 348.

 $^{1}$ H NMR (DMSO–d<sub>6</sub>) δ ppm: 12.62 (s, 1H, OH–5), 8.07 (d, J=8.9 Hz, 2H, H–2', H–6'), 6.87 (d, J=8.9 Hz, 2H, H–3', H–5'), 6.43 (d, J=2.0 Hz, 1H, H–8), 6.19 (d, J=2,0 Hz, 1H, H–6), 5.33 (d, J=5.1 Hz, 1H, H–1''), 3.76 – 3.17 (m, 5H of arabinose + H<sub>2</sub>O).

<sup>13</sup>C NMR (DMSO-d<sub>6</sub>) δ ppm: 177.58 (C-4), 164.29 (C-7), 161.24 (C-5), 160.09 (C-4'), 156.38 (C-9), 156.25 (C-2), 133.58 (C-3), 131.03 (C-2', 6'), 120.71 (C-1'), 115.32 (C-3', 5'), 103.97 (C-10), 101.25 (C-1''), 98.76 (C-6), 93.73 (C-8), 71.60 (C-2''), 70.82 (C-3''), 66.06 (C-4''), 64.26 (C-5'').

Acid hydrolysis: kaempferol, arabinose

Kaempferol 3–0– $\beta$ –D (6"–E–p–coumaro-yl)–glucopyranoside (tiliroside) (V), pale yellow needles (70% MeOH), m.p. 263–266°C.

Rf: PC, S-1, 0.90; S-2, 0.32. UV  $\lambda$  max: a) 267, (302), 315, (360); b) 275, (312), 370; c) 275, 308, (322), 398; d) 276, 307, (322), 397; e) 276, (298), 313, 370; f) 268, (302), 316, (360).

IR v max (KBr) 1680 (ester C=O), 1650 cm<sup>-1</sup> ( $\gamma$ -pyron C=O).

<sup>1</sup>H NMR (DMSO–d<sub>6</sub>) δ ppm: 12.57 (s, 1H, OH–5), 7.98 (d, J=8.7 Hz, 2H, H– 2', 6'), 7.36 (d, J=8.7 Hz, 2H, H–2''', 6'''), 7.33 (d, J=16 Hz, 1H, H–7'''-β), 6.85 (d, J=8.7 Hz, 2H, H–3', 5'), 6.78 (d, J=8.7 Hz, 2H, H–3''', 5'''), 6.38 (d, J=2.0 Hz, 1H, H–8), 6.14 (d, J=2.0 Hz, 1H, H–6), 6.10 (d, J=15.9 Hz, 1H, H–8'''-α), 5.44 (d, J=7.5 Hz, 1H, H–1'').

<sup>13</sup>C NMR (DMSO-d<sub>6</sub>) δ ppm: 177.43 (C-4), 166.20 (C-9'''), 164.19 (C-7), 161.17 (C-5), 160.01 (C-4'), 159.81 (C-4'''), 156.46 (C-2), 156.38 (C-9), 144.62 (C-7'''-β), 133.07 (C-3), 130.85 (C-2', 6'), 130.18 (C-2''', 6'''), 124.94 (C-1'''), 129.79 (C-1'), 115.78 (C-3''', 5'''), 115.11 (C-3', 5'), 113.65 (C-8'''-α), 103.89 (C-10), 100.97 (C-1''), 98.79 (C-6), 93.69 (C-8), 76.23 (C-3''), 74.25 (C-2'''), 74.14 (C-5''), 69.98 (C-4''), 62.98 (C-6'').

Acid hydrolysis: kaempferol, glucose and p-coumaric acid; (PC – S-4) red color after visualization with a mixture of 0.5% diazotized sulphanilic acid and 10% Na<sub>2</sub>CO<sub>3</sub>.

**Methyl gallate** (VI), amorphous, white powder (MeOH), m.p. 195–199°C.

Rf: PC S-1 – 0.77; S-2 – 0.70, violet-brown: under UV; dark-blue after spraying 2% FeCl<sub>3</sub>. UV  $\lambda$  max (MeOH): 220, 276 nm.

IR (KBr)  $v \text{ max} - 1691.8 \text{ cm}^{-1}(\text{C=O, ester})$ . <sup>1</sup>H NMR (CD<sub>3</sub>OD)  $\delta$  ppm; 7.03 (s, 2H), 3.81 (s, 3H – Me ester).

#### RESULTS AND DISCUSSION

Five flavonoid compounds (I - V) and methyl gallate (VI) were isolated from the leaves of *Rubus idaeus L*. subspecies culture of Norna using a multistep chromatography technique.

Flavonoids I and II are derivatives of quercetin; their sugar part is composed of glucose (I) and galactose (II) as was evident from the chromatographic analysis of acid hydrolysate; both sugars were connected to C-3 of the aglycone as followed from the UV spectra. The <sup>1</sup>H NMR spectra of these compounds showed in the aglycone region signals typical for quercetin as well as the presence of the anomeric protons of glucose (I) and galactose (II) with the coupling constants characteristic for β-configuration (2,3). Compound I was identified as quercetin 3–0–β–D–glucopyranoside, and compound II as quercetin 3–0– $\beta$ –D–galactopyranoside. Flavonoids III and IV were derivatives of kaemferol: their sugar part was composed of galactose (III) and arabinose (IV), both sugars were connected to C-3 of the aglycone as followed from the UV spectra. H NMR spectrum of III confirmed that it was kaempferol 3–0– $\beta$ –D–galactopyranoside. <sup>13</sup>C NMR spectrum of IV was characteristic in the sugar part for α-L-arabinopyranosides (4). Compound IV was kaempferol 3-0-α-L-arabinopyranoside.

The  $^1H$  and  $^{13}C$  NMR spectra of compound V were in accordance with tiliroside kaempferol 3–0– $\beta$ –D (6''–E–p–coumaroyl)–glucoside. The  $^1H$  NMR spectrum showed a doublet for the anomeric proton of glucose with a large coupling constant, which revealed that glucose was  $\beta$ –linked. Two doublets at 7.33 and 6.10 ppm with the coupling constant J=16 Hz indicated trans configuration of p–coumaric acid.

The <sup>13</sup>C NMR spectrum confirmed that p-coumaric acid was attached to C-6' of glucose. Two dimensional (HMQC) spectrum let us precisely design signals in <sup>1</sup>H and <sup>13</sup>C NMR spectrum. Tiliroside has been described as a chemical compound from the plant family *Tiliaceae* and *Malvaceae*, as well a component of some species of genus *Rubus* (5–8).

<sup>1</sup>H NMR spectrum of **VI** showed two signals; two proton singlets at 7.03 ppm and three proton singlet at 3.81 ppm charecteristic of gallic acid and its methyl ester (3). Compound **VI** was recognized as a methyl gallate.

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Received: 21.11.2002